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54 **Human glycosyltransferase gene.**

57 A human GnT-III gene having a specific restriction map represented by Fig. 1 of the attached Drawings and a length of approximately 1.6 kb is disclosed, together with a human GnT-III gene which is hybridizable with said gene. A process for producing human GnT-III is provided, which comprises incubating a transformant wherein one of the above-mentioned genes is used and harvesting. The sequencing listing shows a part of the base sequence.

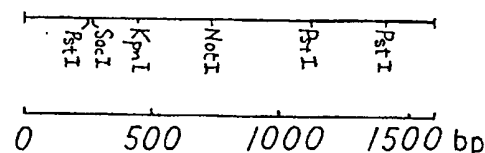


Figure 1

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and examining the capability of the transformant to express human GnT-III. Examples of usable expression cells include COS-1 cells (ATCC CRL 1650). For example, the COS-1 cells can be transformed by the above expression plasmid EX20F. Then the transformant is incubated and the activity of GnT-III expressed in the transformant is determined to specify a gene coding for human GnT-III. This gene is integrated into EX20F and a part of its base sequence is located on a DNA fragment represented by SEQ ID NO. 1. Human GnT-III can be produced by genetic engineering technique by incubating the above transformant.

By effecting hybridization with the use of the gene thus obtained as a probe under strict conditions, it is anticipated that genes for enzymes analogous to that of the present invention, which are different therefrom in sequence but expected to have a similar activity, may be obtained. The term "under stringent conditions" as used herein means that the hybridization of a nylon membrane having DNAs immobilized thereon with the probe is conducted in a solution containing 6 x SSC (1 x SSC means a solution prepared by dissolving 8.76 g of sodium chloride and 4.41 g of sodium citrate in 1 liter of water), 1% of sodium lauryl sulfate, 100 µg/ml of salmon sperm DNA, and 5 x Denhardt's (containing bovine serum albumin, polyvinylpyrrolidone and Ficoll each at a concentration of 0.1%) at 65°C for 20 hours.

As described above in detail, the present invention enables a gene coding for human GnT-III to be isolated and provides a process for producing human GnT-III by using the gene. This gene and its decomposition products are usable in the determination of human GnT-III during the expression process *in vivo* and, therefore, useful in the genetic diagnosis of cancer, and so forth. In addition, various antibodies can be immunologically prepared by using polypeptides coded for by the gene of the present invention. These antibodies are also useful in the field of diagnosis and for the purification of human GnT-III.

Brief Description of the Drawings

Figure 1 is a drawing showing a restriction map of a gene coding for human GnT-III. Figure 2 is a drawing showing relationships among four DNAs H2, H3, H15 and H20. Figure 3 is a drawing showing the construction of a plasmid EX20F.

Example

To further illustrate the present invention in greater detail, and not by way of limitation, the following Examples will be given.

Example 1

(1) Screening of cDNA library

SV3 was prepared from *Escherichia coli* XL1-Blue SV3 (FERM BP-4325) transformed by a plasmid SV3 and the plasmid was digested with *Hind*III to give a DNA fragment of approximately 1.4 kb. This DNA fragment was radiolabeled with [α -³²P] dCTP (3000 Ci/mmol, Amersham) by using a Multiprime DNA Labeling System (Amersham) to thereby give a probe. By using the obtained probe, a human cDNA library [Human Fetal Liver <λgt10>, Clontech] was screened for the target clone by plaque hybridization. As a result, two positive clones were obtained from 3 x 10⁶ plaques. From these clones, DNAs were extracted and digested with *Eco*RI. The digestion products thus obtained were subcloned into Bluescript IISK⁺ and the DNAs thus subcloned (approximately 1.3 kb and approximately 1.5 kb) were respectively named H2 and H3, while the plasmids were respectively named pBluescript II (H2) and pBluescript II (H3). Figure 2 shows the restriction maps of these DNAs and a relationship between them.

(2) Cloning of upstream region containing initiator codon

H2 and H3 were radiolabeled in the same manner as the one described in the above Example 1-(1) to thereby give probes. By using these probes, screening of a human cDNA library was carried out in the same manner as the one described in the above Example 1-(1) to obtain four positive clones from 7 x 10⁵ plaques. The *Eco*RI-digestion products thereof were subcloned into Bluescript IISK⁺. The base sequences of the DNAs thus subcloned were identified and two DNAs containing an initiator codon (approximately 1.6 kb and approximately 1.5 kb) were named respectively H15 and H20, while the plasmids corresponding thereto were named respectively pBluescript II (H15) and pBluescript II (H20). Figure 2 shows the restriction maps of these DNAs and a relationship between thereof with H2 and H3.

Example 2

(1) Construction of expression plasmid

Sequence Listing

5

SEQ ID NO:1

LENGTH:2247

10

TYPE:nucleic acid

STRANDEDNESS:double

TOPOLOGY:linear

15

MOLECULE TYPE:cDNA to mRNA

SEQUENCE DESCRIPTION:SEQ ID NO:1:

20

CCGGCTGCGA TGCCGGGCGC CCGCCGCAGC CGCTGCCGCC GGAGCCCGGG ATGGGGCGAG 60

AGGCTGCGGC GGACGCCAGC ATCTCCCCGC CGGGGACCCC GGGGGCCGCG GAGCCGCCGC 120

CGCCGCTGCT GCGGCCGTTG CTGAGACCCA GCGGGCGATG GGATGAAGAT GAGACGCTAC 180

25

AAGCTCTTTC TCATGTTCTG TATGGCCGGC CTGTGCCTCA TCTCCTTCTT GCACTTCTTC 240

AAGACCCTGT CCTATGTCAC CTCCCCCGA GAACTGGCCT CCCTCAGCCC TAACCTGGTG 300

TCCAGCTTTT TCTGGAACAA TGCCCCGGTC ACGCCCGAGG CCAGCCCCGA GCCAGGAGGC 360

30

CCTGACCTGC TCCGTACCCC ACTCTACTCC CACTCGCCCC TGCTGCAGCC GCTGCCGCCC 420

AGCAAGGCGG CCGAGGAGCT CCACCGGGTG GACTTGGTGC TGCCCGAGGA CACCACCGAG 480

TATTTGCTGC GCACCAAGGC CGGCGGCGTC TGCTTCAAAC CCGGCACCAA GATGCTGGAG 540

35

AGGCCGCCCC CGGGACGGCC GGAGGAGAAG CCTGAGGGGG CCAACGGCTC CTCGGCCCGG 600

CGGCCACCCC GGTACCTCCT GAGCGCCCGG GAGCGCACGG GGGGCCGAGG CGCCCGGCGC 660

AAGTGGGTGG AGTGGGTGTG CCTGCCCGGC TGGCACGGAC CCAGCTGCGG CGTGCCCACT 720

40

GTGGTGCAGT ACTCCAACCT GCCCACCAGG GAGCGGCTGG TGCCCAAGGA GGTGCCGCGC 780

CGCGTCATCA ACGCCATCAA CGTCAACCAC GAGTTCGACC TGCTGGACGT GCGCTTCCAC 840

45

GAGCTGGGCG ACGTGGTGGG CGCCTTTGTG GTGTGCGAGT CCAACTTCAC GGCTTATGGG 900

GAGCCGCGGC CGCTCAAGTT CCGGGAGATG CTGACCAATG GCACCTTCGA GTACATCCGC 960

CACAAGGTGC TCTATGTCTT CCTGGACCAC TTCCCGCCCC GCGGCCGGCA GGACGGCTGG 1020

50

ATCGCCGACG ACTACCTGCG CACCTTCCTC ACCCAGGACG GCGTCTCGCG GCTGCCCAAC 1080

CTGCGGCCCC ACACGCTCTT CATCATTGAC GATCGCGACG AGATCCCGGC CCGTGACGGC 1140

GTCCTTTTCC TCAAGCTCTA CGATGGCTGG ACCGAGCCCT TCGCCTTCCA CATGCCACCG 1200

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	TGC CTC ATC TCC TTC CTG CAC TTC TTC AAG ACC CTG TCC TAT GTC	90
5	Cys Leu Ile Ser Phe Leu His Phe Phe Lys Thr Leu Ser Tyr Val	
	20 25 30	
	ACC TTC CCC CGA GAA CTG GCC TCC CTC AGC CCT AAC CTG GTG TCC	135
10	Thr Phe Pro Arg Glu Leu Ala Ser Leu Ser Pro Asn Leu Val Ser	
	35 40 45	
	AGC TTT TTC TGG AAC AAT GCC CCG GTC ACG CCC CAG GCC AGC CCC	180
15	Ser Phe Phe Trp Asn Asn Ala Pro Val Thr Pro Gln Ala Ser Pro	
	50 55 60	
	GAG CCA GGA GGC CCT GAC CTG CTG CGT ACC CCA CTC TAC TCC CAC	225
20	Glu Pro Gly Gly Pro Asp Leu Leu Arg Thr Pro Leu Tyr Ser His	
	65 70 75	
	TCG CCC CTG CTG CAG CCG CTG CCG CCC AGC AAG GCG GCC GAG GAG	270
25	Ser Pro Leu Leu Gln Pro Leu Pro Pro Ser Lys Ala Ala Glu Glu	
	80 85 90	
	CTC CAC CGG GTG GAC TTG GTG CTG CCC GAG GAC ACC ACC GAG TAT	315
30	Leu His Arg Val Asp Leu Val Leu Pro Glu Asp Thr Thr Glu Tyr	
	95 100 105	
	TTC GTG CGC ACC AAG GCC GGC GGC GTC TGC TTC AAA CCC GGC ACC	360
35	Phe Val Arg Thr Lys Ala Gly Gly Val Cys Phe Lys Pro Gly Thr	
	110 115 120	
	AAG ATG CTG GAG AGG CCG CCC CCG GGA CGG CCG GAG GAG AAG CCT	405
40	Lys Met Leu Glu Arg Pro Pro Pro Gly Arg Pro Glu Glu Lys Pro	
	125 130 135	
	GAG GGG GCC AAC GGC TCC TCG GCC CGG CGG CCA CCC CGG TAC CTC	450
45	Glu Gly Ala Asn Gly Ser Ser Ala Arg Arg Pro Pro Arg Tyr Leu	
	140 145 150	
	CTG AGC GCC CGG GAG CGC ACG GGC GGC CGA GGC CCC CGG CGC AAG	495
50	Leu Ser Ala Arg Glu Arg Thr Gly Gly Arg Gly Ala Arg Arg Lys	
55		

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	Arg Leu Arg Asn Leu Arg Pro Asp Asp Val Phe Ile Ile Asp Asp	
5	305 310 315	
	GCG GAC GAG ATC CCG GCC CGT GAC GGC GTC CTT TTC CTC AAG CTC	990
	Ala Asp Glu Ile Pro Ala Arg Asp Gly Val Leu Phe Leu Lys Leu	
10	320 325 330	
	TAC GAT GGC TGG ACC GAG CCC TTC GCC TTC CAC ATG CGC ACC TCG	1035
	Tyr Asp Gly Trp Thr Glu Pro Phe Ala Phe His Met Arg Thr Ser	
15	335 340 345	
	CTC TAC GGC TTC TTC TGG AAG CAG CCG GGC ACC CTG GAG GTG GTG	1080
	Leu Tyr Gly Phe Phe Trp Lys Gln Pro Gly Thr Leu Glu Val Val	
20	350 355 360	
	TCA GGC TGC ACG GTG GAC ATG CTG CAG GCA GTG TAT GGG CTG GAC	1125
25	Ser Gly Cys Thr Val Asp Met Leu Gln Ala Val Tyr Gly Leu Asp	
	365 370 375	
	GGC ATC CGC CTG CGC CGC CGC CAG TAC TAC ACC ATG CCC AAC TTC	1170
30	Gly Ile Arg Leu Arg Arg Arg Gln Tyr Tyr Thr Met Pro Asn Phe	
	380 385 390	
	AGA CAG TAT GAG AAC CGC ACC GGC CAC ATC CTG GTG CAG TGG TCG	1215
35	Arg Gln Tyr Glu Asn Arg Thr Gly His Ile Leu Val Gln Trp Ser	
	395 400 405	
40	CTG GGC AGC CCC CTG CAC TTC GCC GGC TGG CAC TGC TCC TGG TGC	1260
	Leu Gly Ser Pro Leu His Phe Ala Gly Trp His Cys Ser Trp Cys	
	410 415 420	
45	TTC ACG CCC GAG GGC ATC TAC TTC AAG CTC GTG TCC GCC CAG AAT	1305
	Phe Thr Pro Glu Gly Ile Tyr Phe Lys Leu Val Ser Ala Gln Asn	
	425 430 435	
50	GGC GAC TTC CCA CGC TGG GGT GAC TAC GAG GAC AAG CGG GAC CTG	1350
	Gly Asp Phe Pro Arg Trp Gly Asp Tyr Glu Asp Lys Arg Asp Leu	
55	440 445 450	

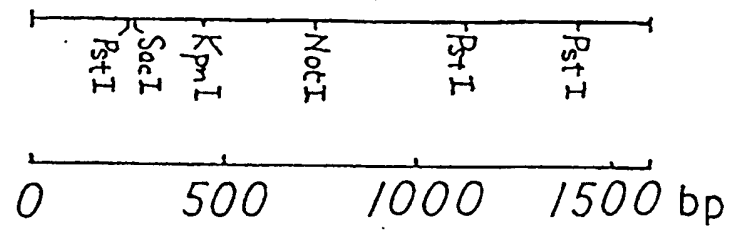


Figure 1

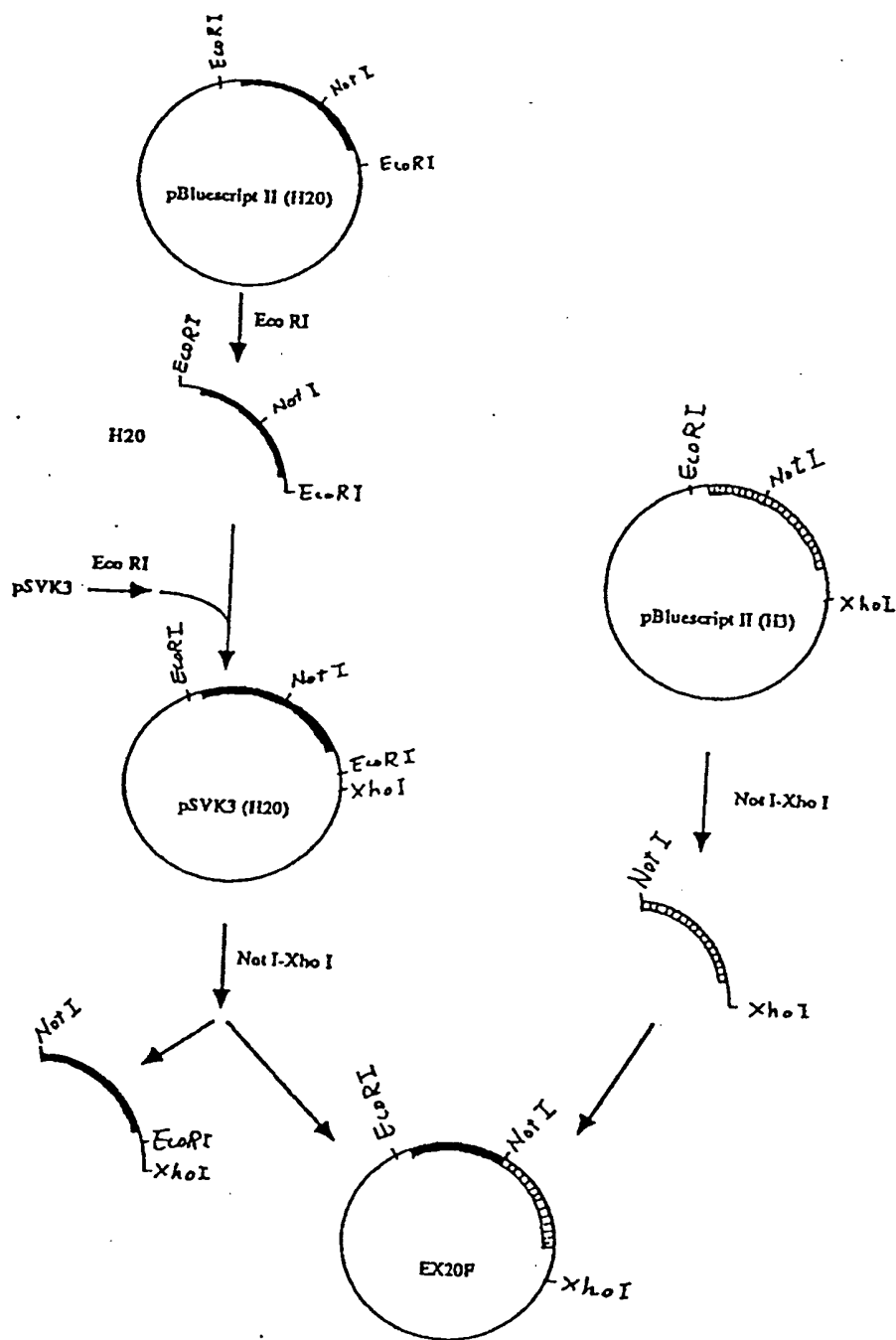


Figure 3